

Phytochemical analysis and docking study of compounds present in a polyherbal preparation used in the treatment of dermatophytosis

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ABSTRACT

Background and Purpose: Soleshine is a polyherbal preparation established in the market for the treatment of cracks and tinea pedis, which is applied externally. This preparation is composed of the extracts of indigenous plants, namely *Azadirachta indica*, *Lawsonia alba*, and *Shorea robusta*, mixed with castor oil and sesame oil. In the present study, an attempt was made to identify the constituents of soleshine and identify some potential drug-like molecules that can inhibit important drug targets of the dermatophytes using molecular docking method.

Materials and Methods: The active ingredients of polyherbal preparation were identified with the aid of gas chromatography-mass spectrometry (GC-MS). Two major compounds were selected based on the retention time and percentage of the area covered in the graph for docking study. The three-dimensional structures of 1,3- β -glucan synthase, chitinase, fungalsin, and lumazine synthase were derived by homology modelling using MODELLER software, version 9.0. The docking of the ligand and receptor was performed using iGEMDOCK and AutodockVina software. The physicochemical properties, lipophilicity, hydrophilicity, and drug likeness properties were obtained from the Swiss ADME online server tool.

Results: The GC-MS analysis demonstrated the presence of different phytochemical compounds in the extract of polyherbal preparation. A total of 20 compounds were identified, among which 3,7-dimethyl-2,6-octadienal and 2-pentene-2-methyl were the major compounds. Regarding 3,7-dimethyl-2,6-octadienal, the covered area and height were 40.15% and 46.17%, respectively. These values were 31.90% and 23.33% for 2-pentene-2-methyl, respectively. These two major compounds had an excellent binding affinity and obeyed the rules for the drug likeness and lead likeness.

Conclusion: As the findings indicated, the two major ingredients present in soleshine showed a good antifungal activity as they inhibited the enzymes responsible for the survival of fungal organism; furthermore, they were appropriate for the lead molecules.

Keywords: Anti-fungal activity, Chitinase, Dermatophytes, Fungalsin, Lumazine synthase, Molecular docking, Soleshine, 1,3- β -Glucan synthase

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Introduction

Diseases caused by fungi mark a vital threat to the health care and are among the critical causes of morbidity and mortality worldwide [1]. In the past, fungi were not considered as important pathogenic organisms as the annual mortality rate due to candidiasis was steady from 1950 to 1970 [2, 3]. However, from 1970 onwards, a significant increase was observed in death rate because of the indiscriminate use of immunosuppressant, broad-spectrum antimicrobial drugs, indwelling intravenous devices, and emergence of viral infections, such as

AIDS. The dreadful consequences of fungal infections has necessitated the search for newer, safer, and more potent drugs [4].

Fungal cell wall is composed of chitin interlinked with 1,3- β -glucan, constituting 30-80% of the cell wall. 1, 3- β -Glucan synthase (EC 2.4.1.34) is an enzyme, which affects the synthesis of fungal cell wall. Chitin, a homopolymer of insoluble linear β 1,4-linked N-acetylglucosamine [5, 6], is a fibrous cellulose-like material, containing chitinases, a poly (1,4- β -[2-acetamido-2-deoxy-D-glucoside]) glycanohydrolases

(EC 3.2.1.14), which are required for the synthesis of the cell wall [7]. Fungalysin, a metallopeptidase, cleaves the proteins and produces metabolic products to enable the activity of exoproteases, resulting in the provision of short peptides and free amino acids for the sustenance of fungal organisms [8].

Lumazine synthase (EC 2.5.1.78) is an enzyme involved in riboflavin (Vitamin B₂) biosynthesis. Bacteria and fungi are not able to incorporate riboflavin exogenously. They absolutely rely on endogenous biosynthesis. Lumazine synthase catalyzes the important steps in riboflavin biosynthesis pathway [9, 10]. All these enzymes are excellent drug targets to prepare new anti-fungal medications.

Soleshine is a polyherbal preparation containing the extracts of the leaves of neem (*Azadirachta indica*) and henna (*Lawsonia alba*), resin of Sal tree (*Shorea robusta*), Sesame oil, and Castor oil. Neem belongs to Meliaceae family, a well-known plant with medicinal properties since olden days. All parts of *Azadirachta indica* have a myriad of medicinal properties [11, 12]. The leaf and bark of the neem plant is used in the treatment of gingivitis, periodontitis, sores, boils, enlarged spleen, malarial fever, fever during childbirth, measles, smallpox, head scald, and cutaneous affections. Neem oil is used as a contraceptive (through intravaginal route for the treatment of vaginal infections), insecticidal agent [13], and mosquito repellent. This oil contains crystalline compounds, namely nimbin and nimbinin, as well as amorphous bitter substance called nimbidin [14].

Henna belongs to the family of Lythraceae, the leaves of which are used in various ailments, such as disarray, jaundice, bleeding disorders, ulcers, prurigo skin diseases, giddiness, and vertigo [15]. The leaves contain naphthoquinones, particularly lawsone, coumarins (laxanthone, I, II, and III), flavonoids, luteolin and its 7-O-glucoside, acacetin-7-O-glucoside, and beta-sitosterol-3-O-glucoside. All parts of the plant contain tannins [16, 17]. The chloroform and ethanolic extracts of henna leaves exhibit promising antimicrobial activity against *Shigella* and *Vibrio cholera*.

Henna plant is used as a prophylactic medicament for the infection of hands and feet against mycosis. The antifungal activity is due to the presence of lawsone, a naphthoquinone that is a known secondary metabolite of henna. The extract of ethanol-water (1:1) of the bark exhibits hepatoprotective activity on carbonate-trichloride-induced liver toxicity. In some experiments with isoplumbagin and lawsaritol, the secondary metabolites obtained from the plant parts showed anti-inflammatory activity.

Shorea robusta, known as a sal tree belongs to the family of Dipterocarpaceae. The bark, young leaves, twigs/leaves, and powder dust of this plant contain 7-12%, 20%, 22%, and 12% tannins, respectively. The aqueous extract of the bark of sal tree contains 39.6% of tannins with a trans/non-trans ratio of 0.73. Oleanolic acid has also been extracted from the bark. Moreover, several triterpenoids have been isolated

from the sal resin [18, 19]. Hydroxyhopanone, dammarenediol II, and dammarenolic acid are reported to be effective as antiviral agents against *Herpes simplex*. The sal resin on dry distillation gives an essential oil, known as Chuua oil containing 96.0% neutral, 3.0% phenolic fraction, and 1.9% acidic fractions. The non-phenolic portion of this oil has an anti-depressant effect on the central nervous system; however, the phenolic portion is less effective.

Sesame oil is obtained from the seeds of *Sesamum indicum* (Family: Pedaliaceae). The oil obtained from sesame seeds is higher in content (around 50%) than that obtained from other seeds [20]. The sesame seeds contain 40-60% oil with almost similar levels of oleic (41%) and linoleic acids (43%) and some palmitic (9%) and stearic acids (6%) [21]. Sesame oil can be classified under oleic-linoleic acid group. The palmitic and stearic [22] are dominant saturated acids. The non-saponifiable fraction of the sesame seed oil entails sterols, lignans, sesamins, nitroslactone, and sesamol. Sesamin and sesamol are not found in any other vegetable oil. Sesamin is present in the concentrations of 0.5% to 1%. Sesamol, a phenolic antioxidant, is present in traces [23].

Castor oil is obtained from *Ricinus communis*, which belongs to the family of Euphorbiaceae. Castor oil obtained from the seeds and young leaves has been traditionally used as laxative and purgative. The gas-liquid chromatography of castor oil showed the availability of ester form of palmitic (1.2%), steric (0.7%), arachidic (0.3%), hexadecenoic (0.2%), oleic (3.2%), linoleic (3.4%), linolenic (0.2%), ricinoleic (89.4%), and dihydroxy stearic acids [24]. The chromatography-mass spectrometry (GC-MS) analysis of castor oil demonstrated the presence of alpha thujone (31.71%) 1,8-cineole (30.98%), alpha-pinene (16.88%), camphor (12.92%), and camphene (7.48%) [25]. Lupeol and 30-norlupan-3 β -ol-20-one were isolated from the coat of castor bean [26].

With this background in mind, the present study aimed to trace out the constituents present in the soleshine using GC-MS analysis and study the inhibition of these compounds against various drug targets of dermatophytes by molecular docking method.

Materials and Methods

Gas chromatography-mass spectrometry

The phytochemical analysis of the extract of soleshine, a polyherbal formulation, was performed by GC-MS equipment (Thermo Scientific Co., Thermo GCTrace ultra, version 5.0, Thermo MS DSQ II). The experimental conditions of GC-MS system included TR 5MS capillary standard non-polar column, dimension of 30 Mts, ID of 0.25 mm, and film thickness of 0.25 μ m. The flow rate of mobile phase (carrier gas: helium) was set at 1.0 mL/min. In the gas chromatography division, the temperature (oven temperature) was 40°C raised to 250°C at 5°C/min, and injection volume was 1 μ l. The samples dissolved in

chloroform were run fully at a range of 50650 m/z, and the results were compared by using Wiley Spectral library search program.

Preparation of protein by homology modeling

The three-dimensional (3D) structures of drug targets selected in this study were not available in the RCSB database. Therefore, their 3D structures were obtained by homology modelling. The primary structures of 1,3- β -glucan synthase (Uniprot accession no: P38631), chitinase (Uniprot accession no: P40954), fungalsin (Uniprot accession no: Q8NIB6), and lumazine synthase (Uniprot accession no: P50861) were obtained in FASTA format from the UniprotKB database.

The homology protein templates of the 1, 3- β -glucan synthase (template RCSB accession code: FKS1), chitinase (template RCSB accession code: 4TX6), fungalsin (template RCSB accession code: 4K90), and lumazine synthase (template RCSB accession code: 1EJB) were obtained from the RCSB database. The homology modeling was performed using MODELLER software (version: 9.0) using EasyModeller as the graphical user interface. The query sequence and template of these proteins were submitted and processed to generate the 3D structures of the proteins. The generated 3D structure of the macromolecule or model protein were validated by means of Ramachandran Plot and SAVES online server tool.

Preparation of ligand

The two compounds, namely 2,6-Octadienal, 3,7-dimethyl and 2-methyl-2-pentene, were selected based on the covered area and height, which were 40.15% and 46.17% in the former compound and 31.90% and 23.33% in the latter one, respectively. The SDF files of these compounds were obtained from Pubchem database. The SDF files were converted into PDB file

format using OPEN BABEL software.

Protein-ligand docking

The initial rough docking was performed in iGEMDOCK software (version 2.0) with a population size of 150 and 70 generations, set as default. Protein-ligand docking was carried out in Autodock Vina [27], which is an interactive molecular graphics program for calculating and displaying the feasible docking modes of protein and ligand pairs presented in a hierarchy based on their binding affinities.

Lead-likeness properties

The SWISS ADME, a free web tool was used to generate the physicochemical, medicinal, and druglikeness properties of these two compounds. Lipinski's rule [28, 29] also called the rule of five (RO5) is to evaluate the druglikeness or determine if a chemical compound with a certain pharmacological or biological activity has properties that may be active peroral.

Toxicity

The toxicity of the compounds was detected with admetSAR, a free online web server. This server provides the possible toxicity profile of the compounds with the values suggesting the safety.

Results

Gas chromatography-mass spectrometry analysis

Figure 1 depicts the thobtained chromatogram. The compounds present in soleshine are demonstrated in Table 1. The chromatogram revealed the presence of 20 compounds in the investigated polyherbal preparation. The two compounds, namely 2,6-Octadienal, 3,7-dimethyl and 2-methyl-2-pentene, were selected for further study on the basis of the covered area and height, which were 40.15% and 46.17% in the former compound and 31.90% and 23.33% in the latter one, respectively.

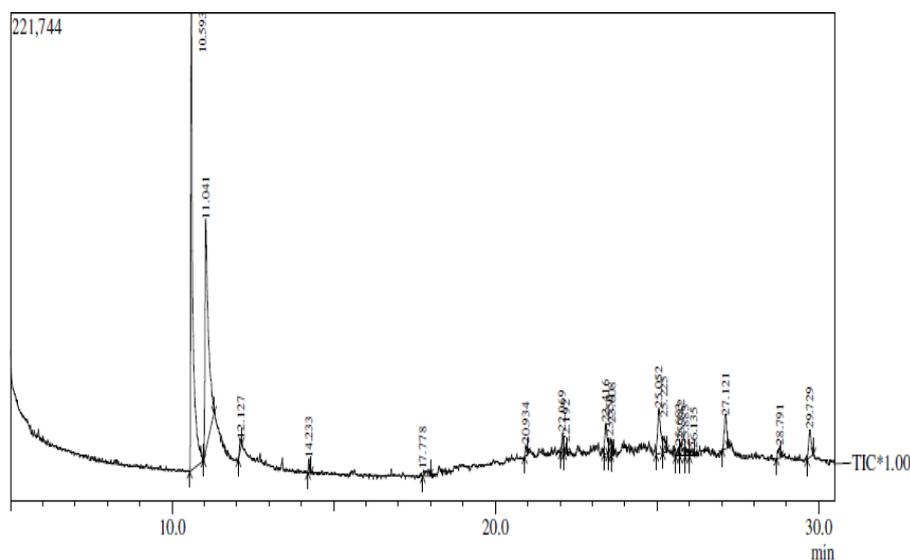
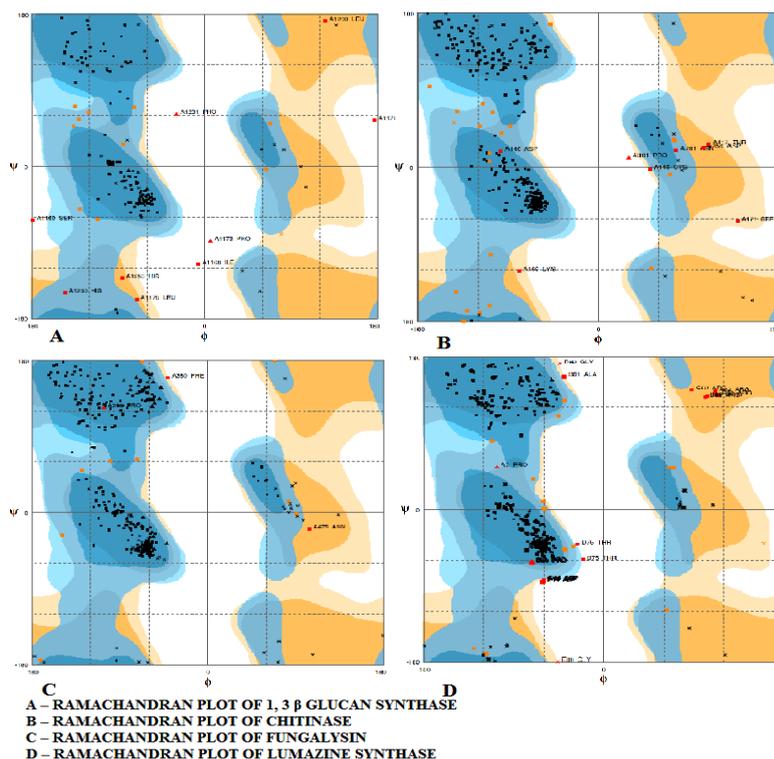


Figure 1. Chromatogram of a polyherbal preparation

Table 1. Compounds obtained from the gas chromatography-mass spectrometry analysis of a poly herbal preparation

Compound No.	R-time	Area%	Height%	Name of the compound	Chemical formula	Mol. weight
1	10.593	40.15	46.17	2,6-Octadienal, 3,7-dimethyl	C ₁₀ H ₁₆ O	152
2	11.041	31.90	23.33	2-Pentene, 2-methyl	C ₆ H ₁₂	84
3	12.127	0.85	0.93	1,2,3-Propanetriol, diacetate	C ₇ H ₁₂ O ₅	176
4	14.233	0.62	1.4 1	Butane, 1,1'-oxybis[4-chloro	C ₈ H ₁₆ ClO	198
5	17.778	1.07	0.62	Oxalic acid, cyclobutyl heptyl ester	C ₁₃ H ₂₂ O ₄	242
6	20.934	0.69	1.13	Docosane	C ₂₂ H ₄₆	310
7	22.069	1.08	1.92	2-Octyldodecan-1-ol	C ₂₀ H ₄₂ O	298
8	22.192	0.64	1.05	3-Methyl-1-(2-tetrahydropyranloxy)-2-butene	C ₉ H ₁₄ O ₃	170
9	23.416	2.84	3.12	Dihexylsulfide	C ₁₂ H ₂₆ S	202
10	23.550	1.29	1.65	Sulfurous acid, cyclohexylmethyl heptadecyl ester	C ₂₄ H ₄₈ O ₃ S	416
11	23.608	0.59	1.4 1	Beta-1-rhamnopyranoside, phenyl-2,3-o-ethylboranediyl-4-o-benzyl	C ₂₁ H ₂₅ BO ₅	368
12	25.052	5.66	4.60	1-(Hexadecyloxy)ethylene	C ₁₈ H ₃₆ O	268
13	25.225	1.38	1.61	2-Butyn-1-al diethyl acetal	C ₈ H ₁₄ O ₂	142
14	25.683	0.71	0.96	5-(Benzyloxy)-7,7-dimethyl-1,3,8-nonatriene	C ₁₈ H ₂₄ O	256
15	25.742	1.51	1.28	1,2,4-Thiadiazol-5(4h)-one, 3-methyl-4-propyl	C ₆ H ₁₀ N ₂ OS	158
16	25.883	0.85	0.89	2,2-Dimethyl-1-propyl phenyl telluride	C ₁₁ H ₁₄ Te	276
17	26.135	0.66	0.73	1-(2-Propenyl)-1-(tosyloxy)cyclopropane	C ₁₃ H ₁₆ O ₃ S	252
18	27.121	3.56	3.37	Dodecane, 1,1'-oxybis-	C ₂₄ H ₅₀ O	354
19	28.791	1.04	1.15	Di-isodecyl phthalate	C ₂₈ H ₄₆ O ₄	446
20	29.729	2.93	2.68	Spiro[cyclopentane-1,2'(1'h)-quinoxaline], 3'-(4-morpholinyl)-6',8'-dinitro	C ₁₆ H ₁₉ N ₅ O ₅	361

R-time: retention time

**Figure 2.** Ramachandran plot of the four proteins showing most of the residues clustered tightly in the most-favoured regions with very few outliers

Homology modeling

The 3D structures obtained by homology modeling were validated with the aid of the Ramachandran plot as shown in Figure 2. The Ramachandran plot demonstrated that most of the residues clustered tightly in the most-favoured regions with very few outliers for all the drug targets. The Ramachandran plot (discovered

by G. N. Ramachandran, C. Ramakrishnan, and V. Sasisekharan [30]) is a way to visualize the dihedral angles, namely ψ (psi) and ϕ (phi), of a protein backbone [31]. Given that steric hindrances occur between adjacent atoms within a protein structure, ψ (psi) and ϕ (phi) values are usually constrained within the specific areas of the plot for ordered structures, such as helices

and sheets. The 1,3- β -glucan synthase protein structure contained 87.5% amino acid residues in the favoured region, 6.9% in the allowed region, and 5.6% in the disallowed region.

In case of chitinase, the favoured, allowed, and disallowed regions were 91.6%, 6.1%, and 2.3%, respectively. The fungalsin was 96.9% favoured, 2.3% allowed, and 0.8% disallowed. Furthermore, lumazine synthase was calculated as 93.8% favoured, 3.6% allowed, and 2.6% disallowed. The proteins were further validated using SAVES server.

Molecular docking

The target proteins, namely 1,3- β -glucan synthase, chitinase, fungalsin, and lumazine synthase, were docked with 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene by iGEMDOCK and Autodock Vina. Clotrimazole,

which is an anti-fungal drug, was also included in the docking study. The energy values and the binding affinities are presented in Table 2. The energy values obtained by iGEMDOCK of the drug targets of 1,3- β -glucan synthase, chitinase, fungalsin, and lumazine synthase, along with 2,6-octadienal, 3,7-dimethyl, 2-methyl-2-pentene, and clotrimazole were -69.94/-65.71/-128.424, -76.36/-83.77/-143.803, -67.49/-67.88/-105.115, and -66.76/-75.84/-115.185 Kcal/mol, respectively.

The binding affinity values obtained by Autodock Vina for 1,3- β -glucan synthase, chitinase, fungalsin, and lumazine synthase, along with 2,6-octadienal, 3,7-dimethyl, 2-methyl-2-pentene, and clotrimazole were -5.8/-5.8/-7.1, -7.2/-7.2/-10.1, -5.1/-5.1/-7.9, and -5.8/-5.8/-8.1, respectively. The docking pose of the compounds with various drug targets were analyzed with LigPlot⁺ software tool. Figures 3-6 displays the

Table 2. Results of rough docking and accurate docking performed with a software iGEMDOCK and Autodock Vina between the drug targets with ligands (2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene) and clotrimazole

S no	Drug targets or protein with ligand	Rough docking energy values with iGEMDOCK					Binding affinity with Autodock Vina		
		Total energy (Kcal/mol)	V.D.W. (Kcal/mol)	H. Bond (Kcal/mol)	Electrostatic (Kcal/mol)	Aver Con pair (Kcal/mol)	Binding affinity	RMSD/UB	RMSD/LB
1	1,3- β -Glucan synthase + 2, 6-octadienal, 3,7-dimethyl	-68.81	-62.55	-6.25	0	34.36	-5.8	0	0
2	1,3- β -Glucan synthase + 2-methyl-2-pentene	-39.50	-39.50	0	0	40.5	-4.2	0	0
3	1,3- β -Glucan synthase + clotrimazole	-128.424	-124.853	-3.5713	0	29.4	-7.1	0	0
4	Chitinase + 2, 6-octadienal, 3,7-dimethyl	-80.59	-75.11	-5.47	0	40.72	-7.2	0	0
5	Chitinase + 2-methyl-2-pentene	-41.99	-41.99	0	0	42.33	-5.4	0	0
6	Chitinase + clotrimazole	-143.803	-140.303	-3.5	0	38	-10.1	0	0
7	Fungalsin + 2, 6-octadienal, 3,7-dimethyl	-64.14	-53.91	-10.22	0	33.27	-5.1	0	0
8	Fungalsin + 2-methyl-2-pentene	-34.21	-34.21	0	0	34.5	-3.7	0	0
9	Fungalsin + clotrimazole	-105.115	-100.119	-4.99543	0	24.08	-7.9	0	0
10	Lumazine synthase + 2,6-octadienal, 3,7-dimethyl	-71.77	-71.77	0	0	40.27	-5.8	0	0
11	Lumazine synthase + 2-methyl-2-pentene	-40.94	-40.94	0	0	39.5	-4.2	0	0
12	Lumazine synthase + clotrimazole	-115.185	-109.823	-5.36169	0	31.44	-8.1	0	0

VDW: Van der Waals force, H Bond: hydrogen bond, RMSD: root mean square deviation, UB: upper bound, LB: lower bound

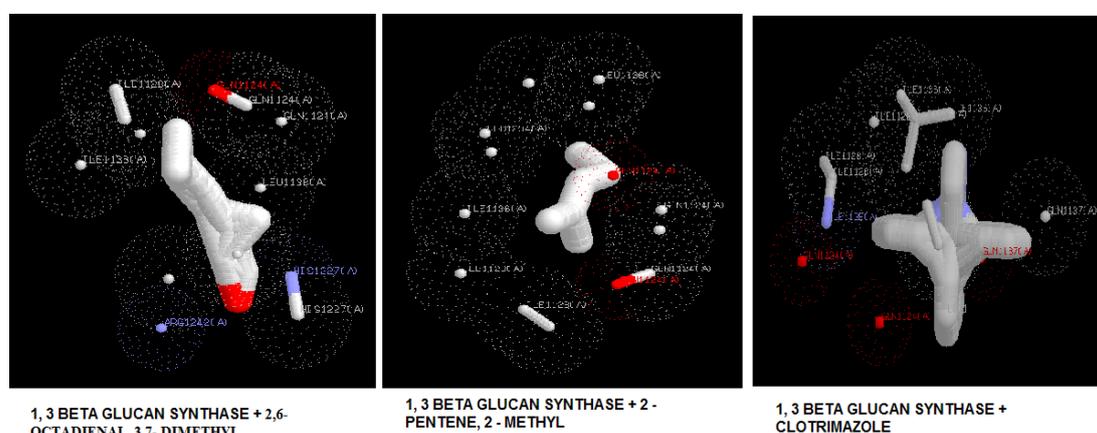


Figure 3. Docking poses of 1,3- β -glucan synthase with 2,6-octadienal, 3,7-dimethyl, 2-methyl-2-pentene, and clotrimazole

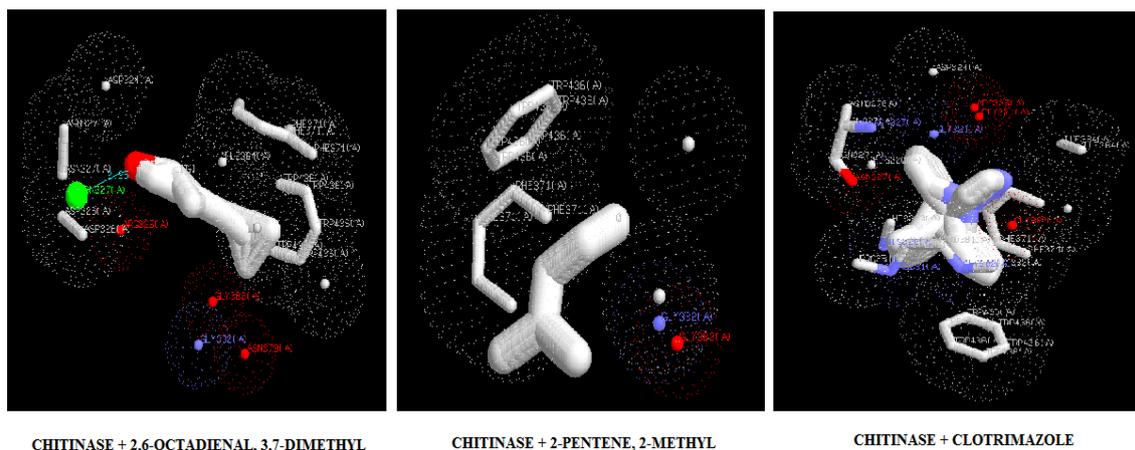


Figure 4. Docking poses of chitinase with 2,6-octadienal, 3,7-dimethyl, 2-methyl-2-pentene, and clotrimazole

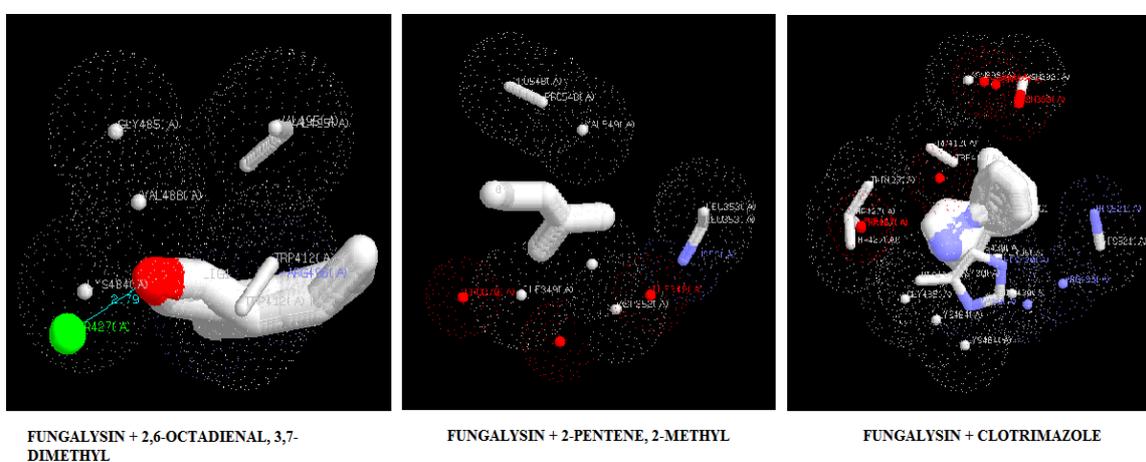


Figure 5. Docking poses of fungalyisin with 2,6-octadienal, 3,7-dimethyl, 2-methyl-2-pentene, and clotrimazole

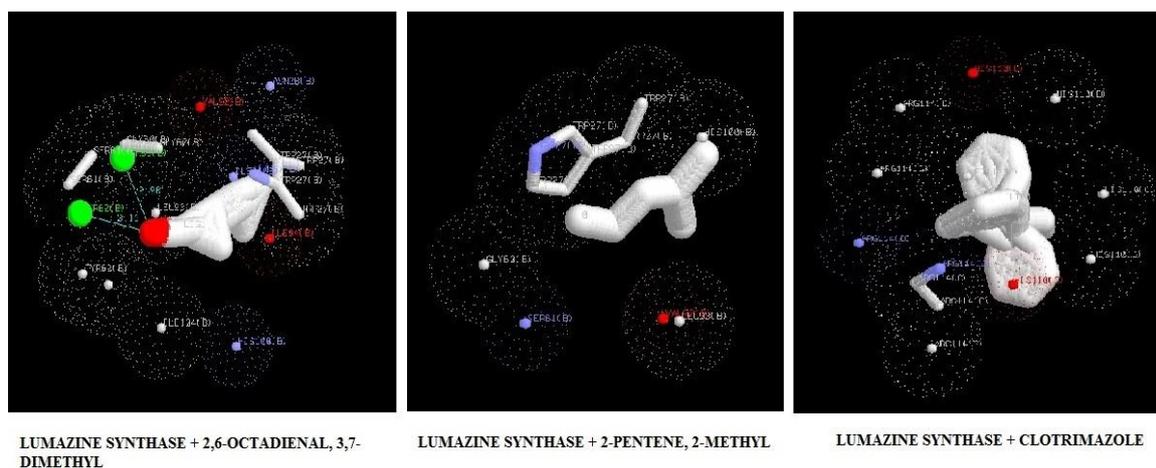


Figure 6. Docking poses of lumazine synthase with 2,6-octadienal, 3,7-dimethyl, 2-methyl-2-pentene, and clotrimazole

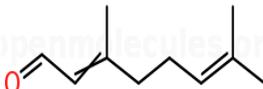
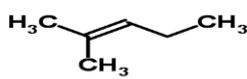
docking poses of various compounds with their protein drug targets. The docking poses were analyzed, and the amino acid residues involved in the various interactions were evaluated

Druglikeness and other properties

Table 3 presents the general properties of

compounds 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene, such as molecular formula, chemical structure, simplified molecular input line entry specification, and international union of pure and applied chemistry name. Table 4 tabulates the molecular weight, number of atoms, fraction CSP3, number of rotatable bonds, molar refractivity, and

Table 3. General properties of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene

Name the of Ligand / compound	Chemical formula	Structure	SMILES	IUPAC Name
2,6-Octadienal, 3,7-dimethyl	C ₁₀ H ₁₆ O		CC(C)=CCCC(C)=CC=O	3,7-dimethylocta-2,6-dienal
2-Methyl-2-pentene	C ₆ H ₁₂		CCC=C(C)C	2-methylpent-2-ene

SMILES: Simplified Molecular Input Line Entry Specification, IUPAC: International Union of Pure and Applied Chemistry

Table 4. Physicochemical properties of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene

Name of ligand	Molecular Weight (g/mol)	Num. heavy atoms	Num. arom. heavy atoms	Fraction CSP3	Num. rotatable bonds	Num. H-bond acceptors	Num. H-bond donors	Molar refractivity	TPSA (Å ²)
2,6-Octadienal, 3,7-dimethyl	152.23	11	0	0.5	4	1	0	49.44	17.07
2-Methyl-2-pentene	84.16	6	0	0.67	1	0	0	30.48	0

TPSA: Topological Polar Surface Area

Table 5. Lipophilicity and hydrophilicity of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene

Name of ligand	Lipophilicity				Hydrophilicity					
	Consensus Log P _{o/w}	Log S (ESOL)	Solubility	Class	Log S (Ali)	Solubility	Class	Log S (SILICOS-IT)	Solubility	Class
2,6-Octadienal, 3,7-dimethyl	2.71	-2.43	5.67E-01	Soluble	-3.05	1.34E-01	Soluble	-1.96	1.66E+00	Soluble
2-Methyl-2-pentene	2.43	-2.01	8.24E-01	Soluble	-2.37	3.56E-01	Soluble	-1.43	3.16E+00	Soluble

o/w: octanol/water

topological polar surface area. The molecular weights, number of atoms, molar refractivity, and polar surface area were less than 500, 20, 50, and 20 Å², respectively, in 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene, representing good oral bioavailability.

Table 5 demonstrates the octanol-water partition coefficient values of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene. As indicated in this table, these values were within the permissible range of -0.4 to +5.6, implying a good lipophilic compound. 2,6-Octadienal, 3,7-dimethyl and 2-methyl-2-pentene compounds were mostly soluble in aqueous medium as the log S was less than -4.0. Table 6 illustrates the pharmacokinetic properties of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene. According to the results, the oral bioavailability was high for 2,6-octadienal, 3,7-dimethyl and low for 2-methyl-2-pentene. Both compounds cross blood brain barrier,

and none of them affected the liver cytochrome P450 enzymes; however, penetration through skin was better for both of the compounds.

Based on Table 7, 2, 6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene followed the Lipinski's rule of 5 [28, 29] and other filters, like Veber [32] and Egan [33], with one violation for Ghose filter [34] of 2,6-octadienal, 3,7-dimethyl, three violations for 2-methyl-2-pentene and two violations of Muegge filter [35]. For a new drug molecule, the bioavailability score [36] is to predict the probability of a new drug that have at least 10% oral bioavailability in rodents. The filters for leadlikeness, like pains filter [37] and brek filter [38], were obeyed for 2, 6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene. A new drug compound with a molecular weight of 250-350, XLog P less than 3.5, rotatable bonds of 7, and synthetic accessibility 2.5 can be an Investigational New Drug (IND).

Table 6. Pharmacokinetics properties of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene

Name of ligand	GI absorption	BBB permeability	P-gp substrate	CYP 1A2 Inhibitor	CYP2C19 Inhibitor	CYP2C9 Inhibitor	CYP2D6 Inhibitor	CYP3A4 Inhibitor	Log K _p (skin permeation) cm/s
2,6-Octadienal, 3,7-dimethyl	High	Yes	No	No	No	No	No	No	-5.08
2-Methyl-2-pentene	Low	Yes	No	No	No	No	No	No	-4.88

GI absorption: Gastrointestinal absorption, BBB: blood brain barrier, CYP: cytochrome P

Table 7. Druglikeness and leadlikeness of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene

Name of ligand	Druglikeness					Leadlikeness				
	Lipinski	Ghose	Veber	Egan	Muegge	Bioavailability score	Pains	Brenk	Leadlikeness	Synthetic accessibility
2,6-Octadienal, 3,7-dimethyl	0	1	0	0	2	0.55	0	3	1	2.49
2-Methyl-2-pentene	0	3	0	0	2	0.55	0	1	1	2.08

Table 8. Toxicity profile of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene

Name of ligand	<i>hERG</i> inhibition	AMES toxicity	Carcinogenicity (Class III)	Acute oral toxicity	Rat acute toxicity (LD 50 mg/)
2,6-Octadienal, 3, 7-dimethyl	0.9220	0.9133	0.5545	0.8232	1.6001
2-Methyl-2-pentene	0.9451	0.9354	0.5328	0.7693	1.6545

hERG: human Ether-a-go-go related gene

Table 8 tabulates the toxicity profile of the compounds of 2,6-octadienal, 3,7-dimethyl and 2-methyl-2-pentene, which were non-toxic in *hERG*, AMES toxicity, acute oral toxicity, and LD50 in rats. Regarding the carcinogenicity, 2,6-octadienal, 3,7-dimethyl was found to be non-carcinogenic, and 2-methyl-2-pentene was revealed to be an alarming sign as the median toxic dose was above 10 mg per kg body weight per day.

Discussion

The GC-MS analysis revealed the presence of many compounds in the polyherbal preparation under investigation. The first major compound, namely 2,6-octadienal, 3,7-dimethyl, showed the retention time, area, and height of 10.593%, 40.15%, and 46.17%, respectively. Regarding the other major compound (i.e., 2-methyl-2-pentene), the retention time, area, and height were 11.041%, 31.90%, and 23.33%, respectively. These two compounds showed good binding affinity with the biological targets of fungal organism, good total energy, and Van der Waals force as depicted in Table 2.

The two compounds present in the polyherbal preparation had a good investigational new drugs as they followed Lipinski's rule of five, and also obeyed druglikeness and leadlikeness properties, as given in tables 5-8. Many studies have been conducted to identify the constituents of the plant extracts having antifungal activities. According to Jeyam et al., 20 phytochemical constituents interacted with 1,3- β -glucan synthase. They showed that the inhibition of 1,3- β -glucan synthase was better by the Echinocandin group of antifungal agents [39].

In another similar study, Mahmoud found that the organic extracts of neem leaves demonstrated more antifungal activity than its aqueous extract [40]. In a study performed by Kannahi, the ethanolic extract of *Lawsonia inermis* showed 100% antifungal activity; however, its aqueous extract demonstrated no activity [41]. According to Laszlo Sami, the inhibition of chitinase by allosamindin showed the growth and survival of fungal organism [42]. In this study, an attempt was made to identify the bioactive compounds of a polyherbal preparation by *in silico* methods.

According to the results, the two compounds showed a good antifungal activity as they inhibited the enzymes responsible for the survival of fungal organism; furthermore, they were appropriate for the lead molecules.

Conclusion

The polyherbal preparation should be further explored to prepare investigational new drugs for the treatment of dermatophytosis. The compounds present in this preparation could be a good target for the proteins that may hamper the survival and growth of the fungi, including dermatophytes.

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Author's contribution

S. N. served as the principal investigator, M. M. supervised the study process, I. K. assisted in the supervision of the study process, and S. V. provided the required software for the study.

Conflicts of interest

The authors of the current study declare no conflicts of interest.

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References

- CSIR. Wealth of India. New Delhi, India: Publications & Information Directory; 1998. P. 164.
- Anaissie EJ, Bodey GP, Rinaldi MG. Emerging fungal pathogens. *Eur J Clin Microbiol Infect Dis.* 1989; 8(4):323-30.
- Wey SB, Mori M, Pfaller MA, Woolson RF, Wenzel RP. Hospital-acquired candidemia. *Arch Intern Med.* 1988; 148:2642-5.
- Beck-Sague C, Banerjee S, Jarvis WR. Infectious diseases and mortality among US nursing home residents.

- Am J Public Health. 1993; 83(12):1739-42.
5. Austinn PR, Brine C J, Castlej E, Zikakisj P. Chitin: New facets of research. *Science*. 1981; 212(4496): 749-53.
 6. Peberdy JF. Fungal cell walls- a review. *Biochemistry of cell walls and membranes in fungi*. Berlin, Heidelberg: Springer; 1990. P. 5-30.
 7. Gooday GW. Biosynthesis of the fungal wall-mechanisms and implications. *J Gen Microbiol*. 1977; 99(1):1-11.
 8. Lemsaddek L, Chambel L, Tenreiro R. Incidence of fungalsyn and subtilisin virulence genes in dermatophytes. Spain: Current Research, Technology and Education Topics in Applied Microbiology and Microbial Biotechnology A; 2010. P. 658-65.
 9. Kearney EB, Goldenberg J, Lipsick J, Perl M. Flavokinase and FAD synthetase from *Bacillus subtilis* specific for reduced flavins. *J Biol Chem*. 1979; 254(19):9551-7.
 10. Gerhardt S, Haase I, Steinbacher S, Kaiser JT, Cushman M, Bacher A, et al. The structural basis of riboflavin binding to *Schizosaccharomyces pombe* 6,7-dimethyl-8-ribityllumazine synthase. *J Mol Biol*. 2002; 318(5): 1317-29.
 11. Brahmachari G. Neem--an omnipotent plant: a retrospection. *Chembiochem*. 2004; 5(4):408-21.
 12. Akhila A, Rani K. Chemistry of the neem tree (*Azadirachta indica* A. Juss.). *Fortschr Chem Org Naturst*. 1999; 78:47-149.
 13. Pandreka A, Dandekar DS, Haldar S, Uttara V, Vijayshree SG, Mulani FA, et al. Triterpenoid profiling and functional characterization of the initial genes involved in isoprenoid biosynthesis in neem (*Azadirachta indica*). *BMC Plant Biol*. 2015; 15(1):214.
 14. Siddiqui S, Siddiqui BS, Faizi S, Mahmood T. Tetracyclic triterpenoids and their derivatives from *Azadirachta indica*. *J Nat Prod*. 1988; 51(1):30-43.
 15. Reddy KR. Folk medicine from Chittoor District, Andhra Pradesh, India used in the treatment of jaundice. *Int J Crude Drug Res*. 1988; 26(3):137-40.
 16. Kawo AH, Kwa AM. Phytochemical screening and antibacterial activity of the aqueous extracts and fractions of ethanolic extracts of *Lawsonia inermis* leaf. *Int Res J Microbiol*. 2011; 2(12):510-6.
 17. Oyedeji AO, Ekundayo O, Koenig WA. Essential oil composition of *Lawsonia inermis* L. leaves from Nigeria. *J Essential Oil Res*. 2005; 17(4):403-4.
 18. Misra LN, Ahmad A. Triterpenoids from *Shorea robusta* resin. *Phytochemistry*. 1997; 45(3):575-8.
 19. Hota RK, Bapuji M. Triterpenoids from the resin of *Shorea robusta*. *Phytochemistry*. 1993; 32(4):466-8.
 20. Hwang LS. *Vegetable Oils* (ed) in *Bailey's Industrial Oil and Fat Products*. 6th ed. New Jersey: John Wiley & Sons; 2005. P. 1178.
 21. Gunstone FD. *The chemistry of oils and fats: sources, composition, properties and uses*. 1st ed. Oxford: Blackwell Publishing; 2004. P. 8.
 22. Nzikou JM, Matos L, Bouanga-Kalou G, Ndangui CB, Pambou-Tobi NP, Kimbonguila A, et al. Chemical composition on the seeds and oil of sesame (*Sesamum indicum* L.) grown in Congo-Brazzaville. *Adv J Food Sci Technol*. 2009; 1(1):6-11.
 23. Khare CP. *Indian medicinal plants: an illustrated dictionary*. New York: Springer Science & Business Media; 2007.
 24. Kang SS, Cordell GA, Soejarto DD, Fong HHS. Alkaloids and flavonoids from *Ricinus communis*. *J Nat Prod*. 1985; 48(1):155-6.
 25. Kadri A, Gharsallah N, Damak M, Gdoura R. Chemical composition and in vitro antioxidant properties of essential oil of *Ricinus communis* L. *J Med Plants Res*. 2011; 5(8):1466-70.
 26. Thompson MJ, Bowers WS. Lupeol and 30-norlupan-3 β -ol-20-one from the coating of the castor bean (*Ricinus communis* L.). *Phytochemistry*. 1968; 7(5):845-7.
 27. Trott O, Olson AJ. AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading. *J Comput Chem*. 2010; 31(2):455-61.
 28. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev*. 2001; 46(1-3):3-26.
 29. Lipinski CA. Lead and drug like compounds: the rule of five revolution. *Drug Discov Today Technol*. 2004; 1(4):337-41.
 30. Richardson JS. Anatomy and taxonomy of protein structures. *Adv Protein Chem*. 1981; 34:167-339.
 31. Ramachandran GN, Ramakrishnan C, Sasisekharan V. Stereochemistry of polypeptide chain configurations. *J Mol Biol*. 1963; 7(1):95-9.
 32. Veber DF, Johnson SR, Cheng HY, Smith BR, Ward KW, Kopple KD. Molecular properties that influence the oral bioavailability of drug candidates. *J Med Chem*. 2002; 45(12):2615-23.
 33. Egan WJ, Merz KM, Baldwin JJ. Prediction of drug absorption using multivariate statistics. *J Med Chem*. 2000; 43(12):3867-77.
 34. Ghose AK, Viswanadhan VN, Wendoloski JJ. A knowledge-based approach in designing combinatorial or medicinal chemistry libraries for drug discovery. 1. A qualitative and quantitative characterization of known drug databases. *J Comb Chem*. 1999; 1(1): 55-68.
 35. Muegge I, Heald SL, Brittelli D. Simple selection criteria for drug-like chemical matter. *J Med Chem*. 2001; 44(12):1841-6.
 36. Martin YC. A bioavailability score. *J Med Chem*. 2005; 48(9):3164-70.
 37. Baell JB, Holloway GA. New substructure filters for removal of pan assay interference compounds (PAINS) from screening libraries and for their exclusion in bioassays. *J Med Chem*. 2010; 53(7):2719-40.
 38. Brenk R, Schipani A, James D, Krasowski A, Gilbert IH, Frearson J, et al. Lessons learnt from assembling screening libraries for drug discovery for neglected diseases. *Chem Med Chem*. 2008; 3(3):435-4.
 39. Jeyam M, Arangaraj M, Ravikumar P, Shalini G. Computational analysis of phytocompounds with 1, 3 - β -D-Glucan synthase for antidermatophytic activity. *J App Pharm Sci*. 2014; 4(2):64-9.
 40. Mahmoud DA, Hassanein NM, Youssef KA, Zeid MA. Antifungal activity of different neem leaf extracts and the nimonol against some important human pathogens. *Brazil J Microbiol*. 2011; 42(3):1007-16.
 41. Mannargudi TD. Antimicrobial activity of *Lawsonia inermis* leaf extracts against some human pathogens. *Int J Curr Microbiol Appl Sci*. 2013; 2(5):342-9.
 42. Sámi L, Pusztahelyi T, Emri T, Varcza Z, Fekete A, Grallert Á, et al. Autolysis and aging of *Penicillium chrysogenum* cultures under carbon starvation: Chitinase production and antifungal effect of allosamidin. *J Gen Appl Microbiol*. 2001; 47(4):201-11.