

## *Pneumocystis jirovecii* colonization in Chronic Obstructive Pulmonary Disease (COPD)

Khodavaisy S<sup>1,2</sup>, Mortaz E<sup>3</sup>, Mohammadi F<sup>2</sup>, Aliyali M<sup>4</sup>, Fakhim H<sup>5</sup>, Badali H<sup>6\*</sup>

<sup>1</sup> Department of Medical Parasitology and Mycology, Kurdistan University of Medical Sciences, Sanandaj, Iran

<sup>2</sup> Department of Medical Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

<sup>3</sup> Division of Pharmacology, Utrecht Institute for Pharmaceutical Sciences, Faculty of Science, Utrecht University, Utrecht, The Netherlands

<sup>4</sup> Pulmonary and Critical Care Division, Mazandaran University of Medical Sciences, Sari, Iran

<sup>5</sup> Student Research Committee, Mazandaran University of Medical Sciences, Sari, Iran

<sup>6</sup> Department of Medical Mycology and Parasitology/Invasive Fungi Research Center, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran

\*Corresponding author: Hamid Badali, Department of Medical Mycology & Parasitology, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran; Post Code: 48175-1665. Tel: +98 11 33543781; Fax: +98 11 33543781; Email: badalii@yahoo.com

(Received: 27 September 2014; Revised: 11 November 2014; Accepted: 3 January 2015)

### Abstract

Chronic obstructive pulmonary disease (COPD) is associated with a chronic inflammatory response in airways and lung parenchyma that results in significant morbidity and mortality worldwide. Cigarette smoking considered as an important risk factor plays a role in pathogenesis of disease. *Pneumocystis jirovecii* is an atypical opportunistic fungus that causes pneumonia in immunosuppressed host, although the low levels of its DNA in patients without signs and symptoms of pneumonia, which likely represents colonization. The increased prevalence of *P. jirovecii* colonization in COPD patients has led to an interest in understanding its role in the disease. *P. jirovecii* colonization in these patients could represent a problem for public health since colonized patients could act as a major reservoir and source of infection for susceptible subjects. Using sensitive molecular techniques, low levels of *P. jirovecii* DNA have been detected in the respiratory tract of certain individuals. It is necessary to elucidate the role of *P. jirovecii* colonization in the natural history of COPD patients in order to improve the clinical management of this disease. In the current review paper, we discuss *P. jirovecii* colonization in COPD patients..

**Keywords:** *Chronic Obstructive, Pneumocystis jirovecii, Pulmonary Disease, Smoking*

➤ How to cite this paper:

Khodavaisy S, Mortaz E, Mohammadi F, Aliyali M, Fakhim H, Badali H. *Pneumocystis jirovecii* colonization in Chronic Obstructive Pulmonary Disease (COPD). Curr Med Mycol. 2015; 1(1): 42-48. DOI: [10.18869/acadpub.cmm.1.1.42](https://doi.org/10.18869/acadpub.cmm.1.1.42)

### Introduction

Chronic obstructive pulmonary disease (COPD) is a slowly progressive lung diseases characterized by airflow limitation associated with a chronic inflammatory response in both airways and lung parenchyma. It is a major cause of illness and results in significant morbidity and mortality [1]. Nowadays, COPD is the fourth leading cause of mortality among individuals, and estimated to rank third by 2020 around the world [2]. Smoking is considered as a major risk factor for the development of COPD, but not all smokers develop the disease and factors determining the severity or pattern of disease in smokers are largely unknown [3]. As a cause of systemic inflammatory response (increased levels of several circulating cytokines and acute-phase reactants, i.e., IL-8, IL-6 and TNF- $\alpha$ ), infectious agents such as bacteria, viruses

and fungi could therefore play a role in the pathophysiology of COPD [4-5]. Oxidative stress and cells likes neutrophils, macrophages and mast cells playing important role in the pathogenesis of disease. *Pneumocystis jirovecii* is an atypical opportunistic fungus with lung tropism and worldwide distribution that causes pneumonia in immunosuppressed individuals such as HIV [6-7]. *P. jirovecii* colonization has been described in individuals with various lung diseases that may be important in COPD pathogenesis. Morris *et al*, 2008 believed that *P. jirovecii* colonization alone or with tobacco could be a cofactor that increase or maintain inflammatory response, thus developing COPD progression [1].

The increased prevalence of *P. jirovecii* colonization in those with COPD has led to an interest in understanding its role in the disease.

The study in a non-HIV-infected population have demonstrated that *P. jirovecii* colonization is a risk factor for severe COPD (Global Health Initiative on Obstructive Lung Disease [GOLD] Stage IV), independent of smoking history or corticosteroid abuser [8]. Thus, it is more common in subjects with severe COPD than in those with milder or no disease and is associated with a more rapid progression of disease. *P. jirovecii* colonization in COPD patients could represent a problem for public health since colonized patients could act as a major reservoir and source of infection for susceptible subjects [9]. Understanding the role of this colonization in COPD patients much has been discovered about the biology of *P. jirovecii* in understanding the role of this colonization in COPD patients. Therefore in the current review paper, we focus on *P. jirovecii* colonization with emphasis to the epidemiology, diagnosis, and treatment aspects of COPD patients.

**Epidemiology**

Pneumocystis pneumonia (PCP) is a potentially life-threatening infection which is the most common opportunistic infection in immunocompromised individuals and plays a role in the development of airway obstruction. Calderon et al. reported 10% of patients with chronic bronchial disease were colonized with *P. jirovecii* based on staining methods of sputum [10]. In addition in 2004, they revealed *P. jirovecii* colonization in 41% of

patients with chronic bronchitis and COPD by using nested PCR technique [11]. Later, Probst et al, found that more than 21% were colonized with *P. jirovecii* in COPD by using nested PCR of various respiratory samples [12]. *P. jirovecii* colonization rate was reported, 36.7 % in the among smokers group with very severe COPD based on PCR, compared with 5.3 % of smokers with normal spirometry, mild COPD, moderate COPD and severe COPD (p= 0.004) and with 9.1% of control subjects (p= 0.007) [8].

Calderon et al. investigated 51 patients with COPD and reported 28 (55%) were colonized by *P. jirovecii* pneumonia which showed a higher level of proinflammatory cytokines than COPD without PCP [13]. Calderon *et al.*, demonstrated a strong association between *P. jirovecii* colonization and the severity of COPD. This relationship is independent of the smoking history [11]. Another study found that 16% (8/50) of COPD had *P. jirovecii* colonization by performing PCR on sputum specimens [12]. Morris et al. investigated antibodies *P. jirovecii* endoprotease kexin (*anti-KEX1* antibody) in 96 patients with COPD (62.7%) whose smoking backgrounds with at least 10 packs per year. They believed that low or undetectable *anti-KEX1* PCP titer among smokers might increase susceptibility to colonization with *P. jirovecii* and progression of COPD [1]. The prevalence of *P. jirovecii* colonization among COPD patients is different in published studies (Table 1).

**Table 1.** *P. jirovecii* colonization in COPD patients

Patients, no.	Diagnostic Sample	Diagnostic Methods	Population	% Colonization	References
50	sputum	IHC stains, IF stains	Patients with chronic bronchial disease	10.0%	[10]
8	BAL	Nested PCR	Patients with COPD	37.5%	[14]
37	BAL and sputum	Nested PCR	Patients with COPD	41%	[12]
23	BAL, sputum, and tracheal aspirates	Touch-down PCR	In-patients with suspected bacterial pneumonia and COPD	43.5%	[15]
23	BAL	Nested PCR	Patients with COPD undergoing bronchoscopy	17.5%	[16]
37	sputum	Nested PCR	Patients with chronic bronchitis	40.5%	[11]
51	Sputum	Nested PCR	COPD	54.9%	[13]
68	Lung resection	Nested PCR	COPD and other lung diseases	19.1%	[8]
50	Sputum	Nested PCR	COPD	16.0%	[17]

Abbreviation: IHC, immunohistochemical; IF, immunofluorescence; COPD, chronic obstructive pulmonary disease; PCR, polymerase chain reaction

## Diagnosis

Since the diagnosis of uncultivable *P. jirovecii* is a big challenge and definitively confirmed by microscopic identification of the causative organism in sputum or bronchio-alveolar lavage by staining methods (Gomori methenamine (GMS) silver, toluidine blue-O, Giemsa staining, or Diff-Quik), or an immuno-fluorescence assay will show the characteristic cysts [18]. Monoclonal antibodies can be used to detect PCP with a rapid, sensitive, and easy-to-perform immunofluorescence assay [19-20]. These methods are generally not adequate for detection of *P. jirovecii* colonization, and sensitive techniques is highly recommended. Fortunately with the development of diagnostic techniques, diagnoses are now established by less invasive methods and more sensitive [21-23].

Early studies of Pneumocystis surface moieties revealed a predominant surface glycoprotein, the major surface glycoprotein (Msg) [24]. Msg is encoded by a large gene family consisting of over 100 copies. Molecular examination of Msg genomic localization and expression revealed that the msg gene undergoes extensive genomic rearrangement, resulting in variations in its antigenic properties [25]. Walzer et al. developed an Enzyme-linked immunosorbent assay (ELISA) that showed a promise in diagnostic testing and epidemiologic studies [23-24]. Recent studies have supported the utility of MsgC titers as indicators of acute *P. jirovecii* pneumonia [24]. In this line it has been shown that the *P. jirovecii* protease, kexin (Kex1, Prt1) has been investigated for potential use in serologic studies [25-28]. Several studies with human subjects and experimental animal models suggest that PCP Kex may be a useful target for serologic studies and a potential target for immunologic control of PCP colonization [29]. Recently, the development of molecular detection has been instrumental in advancing the study of PCP colonization.  $\beta$ -

1,3-D-glucan is a polysaccharide that is present in the Pneumocystis cyst wall as well as in the walls of most fungi. It triggers an innate immune response which can be detected in BAL and serum specimens from immunocompromised patients with PCP [30]. Quantitative PCR (qPCR) assays provide a very sensitive test for detecting *P. jirovecii*. Respiratory secretions may contain low copy numbers of *P. jirovecii* in individuals who are free of pulmonary dysfunction and who are either immunologically normal or abnormal. Thus, a positive PCR test does not necessarily imply the PCP is the cause of pulmonary dysfunction. Therefore results need to be confirmed in clinical investigations under standardized protocols for specimen collection and PCR performance [31, 32].

## Treatment

Despite the application of prophylaxis and therapy (trimethoprim/sulfamethoxazole) (Table 2) pneumocystis pneumonia due to *P. jirovecii* remains a life-threatening infection with significant morbidity and mortality especially in HIV-infected patients [33-37]. The second line therapies and alternatives to TMP-SMX are intravenous pentamidine, clindamycin with primaquine, dapsone with trimethoprim, atovaquone, and trimetrexate with folinic acid [38-42]. However, intravenous pentamidine is often preferred; its use is associated with a high rate of significant side effects like nephrotoxicity. Clindamycin with primaquine has excellent activity against *P. jirovecii* and the combination is second-line treatment for PCP in patients who fail treatment with TMP-SMX [43-45]. Selection of an initial anti-Pneumocystis regimen depends on the severity of the patient's illness include mild-moderate disease and moderate-severe disease. Oral TMP-SMX is more effective than other regimens which can be given at a dose of 2 double-strength tablets (TMP 160 mg

**Table 2.** Treatment regimens for *P. jirovecii* colonization in COPD

First choice	Alternatives	Adjunctive corticosteroids
TMP-SMX (15–20 mg/kg TMP and 75–100 mg/kg i.v. per day, divided q6h or q8h)	Clindamycin-primaquine, pentamidine (3–4 mg/kg i.v. per day)	Prednisone (40 mg p.o. b.i.d. 5 days, then 40 mg p.o. q.d. 5 days, then 20 mg p.o. q.d. for 11 days), methylprednisolone i.v. at 75% of prednisone dose, start at time of antibiotic initiation or at least within 72 h

and SMX 800 mg) every 8 h. In form of moderate to severe disease, intravenous therapy is preferred than oral therapy, with a dose of 15 to 20 mg/kg TMP and 75 to 100 mg/kg SMX divided every 6 to 8 h. Although, if a patient with moderate is unable to tolerate TMP-SMX, alternative choices as second-line treatment for PCP would be intravenous pentamidine or Clindamycin- primaquine (600 to 900 mg intravenously every 6 to 8 h) with primaquine (15 to 30 mg base orally daily) [46-49]. The alternative choice for mild disease include Atovaquone (750 mg orally given twice daily), dapson (100 mg orally daily) plus TMP (15 mg/kg/day orally in three divided doses), and oral primaquine (15 to 30 mg/daily) plus oral clindamycin 300 to 450 mg every 6 to 8 h. However studies have shown that prolonged use of mono-therapy prophylaxis for *P. jirovecii* colonization might develop the drug resistancy and decreased antibiotic effectiveness [50-52]. Other agents under investigation include echinocandins. There is interest in the activity of caspofungin against the cyst form of *Pneumocystis*, but there is scant clinical evidence that this drug is useful for treating or preventing human disease. *Pneumocystis* spp, appear to have a biphasic life cycle consisting of an asexual phase and a sexual cycle resulting in formation of cysts. The cysts as the agent of transmission contained abundant  $\beta$ -1,3-D-glucan observed in the mouse model, the echinocandins are able to inhibit  $\beta$ -1,3-D-glucan, and reduction of cyst numbers, therefore increases the survival of mice. Thus, understanding the life cycle of this genus is crucially important to improve the managements of infections [53-54]. Although patients may worsen early during PCP treatment due to a transient inflammatory response to the organism, true treatment failure may also develop. Although drug resistance cannot be directly tested in pneumocystis, similar mutations develop in bacteria after trimethoprim-sulfamethoxazole (SXT) exposure and lead to antibiotic resistance [35]. Despite the theoretical concern for drug resistance SXT remains the treatment of choice even in those with previous sulfa exposure.

## Conclusions

COPD is a major public health problem that causes chronic morbidity and mortality throughout the world. Besides, it has been recently demonstrated that colonized patients with COPD have higher systemic pro-inflammatory cytokine levels such as peripheral lymphocyte counts, IL-6, IL-8, and TNF- $\alpha$ , than non-colonized patients. Recent studies have shown that to pay attention on pneumocystis in the progression of chronic pulmonary diseases. Although, *P. jirovecii* colonization is a risk factor for exacerbation of COPD, it is independent of smoking history or corticosteroid usage. *P. jirovecii* colonization could provokes airflow obstruction, with the pathogen acting as a co-morbidity factor that may stimulate pulmonary inflammation and may play a pathologic role in COPD patients. Several studies suggest a potential role of *P. jirovecii* in the pathophysiology of COPD through inducing inflammatory changes with chronic lung destruction and interacting with other risks factors, i.e., tobacco or pathogenic bacteria. The presence of *P. jirovecii* in the lungs, even at low levels, may stimulate a host inflammatory response that leads to lung damage and may play a crucial role in the progression of COPD. Therefore, further investigations are highly recommended to confirm the role of *P. jirovecii* colonization in the pathogenesis of COPD to improve the clinical management.

## Acknowledgements

The authors would like to thank the School of Medicine, Mazandaran University of Medical Sciences, Sari; Iran for the financial support. Ms. Shahi is gratefully acknowledged for her technical assistance and to Prof. Josef Dumanov for critically reviewing and editing the manuscript.

## Authors' contributions

H.B. and S.K. design and writing the draft version of the article. E.M. and M.A. were contributed in data gathering. The rest have completed the draft and contributed in the preparation of the final article.

## Conflicts of interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

## Financial Disclosure

No financial interests related to the material of this manuscript have been declared.

## References

- Morris A, Scieurba FC, Norris KA. Pneumocystis: a novel pathogen in chronic obstructive pulmonary disease? COPD. 2008; 5(1): 43-51.
- Chapman K, Mannino DM, Soriano J, Vermeire P, Buist A, Thun M, et al. Epidemiology and costs of chronic obstructive pulmonary disease. Eur Respir J. 2006; 27(1):188-207.
- Mortaz E, Folkerts G, Engels F, Nijkamp FP, Redegeld FA. Cigarette smoke suppresses in vitro allergic activation of mouse mast cells. ClinExp Allergy. 2009; 39(5): 679-87.
- MacNee W. Pathogenesis of chronic obstructive pulmonary disease. Proc Am Thorac Soc. 2005; 2(4): 258-66.
- Mortaz E, Folkerts G, Redegeld F. Mast cells and COPD. PulmPharmacolTher. 2011; 24(4): 367-72.
- Masur H, Michelis MA, Greene JB, Onorato I, VandeStouwe RA, Holzman RS, et al. An outbreak of community-acquired *Pneumocystis carinii* pneumonia: initial manifestation of cellular immune dysfunction. N Engl J Med. 1981; 305(24): 1431-8.
- Thomas Jr CF, Limper AH. Pneumocystis pneumonia. N Engl J Med. 2004; 350(24): 2487-98.
- Morris A, Scieurba FC, Lebedeva IP, Githaiga A, Elliott WM, Hogg JC, et al. Association of chronic obstructive pulmonary disease severity and Pneumocystis colonization. Am J Respir Crit Care Med. 2004; 170(4): 408-13.
- Wissmann G, Morilla R, Friaza V, Calderón E, Varela JM. El serhumanocomoreservorio de Pneumocystis. Enferm Infecc Microbiol Clin. 2010; 28(1): 38-43.
- Calderon EJ, Regordan C, Medrano FJ, Ollero M, Varela JM. *Pneumocystis carinii* infection in patients with chronic bronchial disease. Lancet. 1996; 347(9006): 977.
- Calderon E, De la Horra C, Medrano F, López-Suárez A, Montes-Cano M, Respaldiza N, et al. Pneumocystis jirovecii isolates with dihydropteroate synthase mutations in patients with chronic bronchitis. Eur J Clin Microbiol Infect Dis. 2004; 23(7): 545-9.
- Probst M, Ries H, Schmidt-Wieland T, Serr A. Detection of *Pneumocystis carinii* DNA in patients with chronic lung diseases. Eur J Clin Microbiol Infect Dis. 2000; 19(8): 644-5.
- Calderón EJ, Rivero L, Respaldiza N, Morilla R, Montes-Cano MA, Friaza V, et al. Systemic inflammation in patients with chronic obstructive pulmonary disease who are colonized with *Pneumocystis jirovecii*. Clin Infect Dis. 2007; 45(2): 17-9.
- Sing A, Roggenkamp A, Autenrieth IB, Heesemann J. *Pneumocystis carinii* carriage in immunocompetent patients with primary pulmonary disorders as detected by single or nested PCR. J Clin Microbiol. 1999; 37(10): 3409-10.
- Helweg-Larsen J, Jensen JS, Dohn B, Benfield TL, Lundgren B. Detection of *Pneumocystis* DNA in samples from patients suspected of bacterial pneumonia-a case-control study. BMC Infect Dis. 2002; 2(1):28.
- Maskell N, Waine D, Lindley A, Pepperell J, Wakefield A, Miller R, et al. Asymptomatic carriage of *Pneumocystis jirovecii* in subjects undergoing bronchoscopy: a prospective study. Thorax. 2003; 58(7): 594-7.
- Nevez G, Raccurt C, Vincent P, Jounieaux V, De-Cas E. Pulmonary Colonization with *Pneumocystis carinii* in Human Immunodeficiency Virus-Negative Patients: Assessing Risk with Blood CD4+ T Cell Counts. Clin Infect Dis. 1999; 29(5): 1331-2.
- Morris A, Kingsley LA, Groner G, Lebedeva IP, Beard CB, Norris KA. Prevalence and clinical predictors of *Pneumocystis* colonization among HIV-infected men. AIDS. 2004; 18(5): 793-8.
- Tasaka S, Hasegawa N, Kobayashi S, Yamada W, Nishimura T, Takeuchi T, et al. Serum indicators for the diagnosis of pneumocystis pneumonia. Chest. 2007; 131(4): 1173-80.
- Ng V, Virani N, Chaisson R, Yajko D, Sphar H, Cabrian K, et al. Rapid detection of *Pneumocystis carinii* using a direct fluorescent monoclonal antibody stain. J Clin Microbiol. 1990; 28(10): 2228-33.
- Tipirneni R, Daly KR, Jarlsberg LG, Koch JV, Swartzman A, Roth BM, et al. Healthcare worker occupation and immune response to *Pneumocystis jirovecii*. Emerg Infect Dis. 2009; 15(10): 1590.
- Walzer PD, Djawe K, Levin L, Daly KR, Koch J, Kingsley L, et al. Long-term serologic responses to the *Pneumocystis jirovecii* major surface glycoprotein in HIV-positive individuals with and without *P. jirovecii* infection. J Infect Dis. 2009; 199(9): 1335-44.
- Daly KR, Huang L, Morris A, Koch J, Crothers K, Levin L, et al. Antibody Response to *Pneumocystis jirovecii*: Antibody Response to *Pneumocystis jirovecii* Major Surface Glycoprotein. Emerg Infect Dis. 2006; 12(8): 1231-7.
- Daly KR, Koch J, Levin L, Walzer PD. Enzyme-linked immunosorbent assay and serologic responses to *Pneumocystis jirovecii*. Emerg Infect Dis. 2004; 10(5): 848-54.
- Crothers K, Daly KR, Rimland D, Goetz MB, Gibert CL, Butt AA, et al. Decreased serum antibody

- responses to recombinant *Pneumocystis* antigens in HIV-infected and uninfected current smokers. *Clin Vaccine Immunol.* 2011; 18(3): 380-6.
26. Djawe K, Daly KR, Vargas SL, Santolaya ME, Ponce CA, Bustamante R, et al. Seroepidemiological study of *Pneumocystis jirovecii* infection in healthy infants in Chile using recombinant fragments of the P. jirovecii major surface glycoprotein. *Int J Infect Dis.* 2010; 14(12): 1060-6.
  27. Gingo MR, Lucht L, Daly KR, Djawe K, Palella FJ, Abraham AG, et al. Serologic responses to *Pneumocystis* proteins in human immunodeficiency virus patients with and without *Pneumocystis jirovecii* pneumonia. *J Acquir Immune Defic Syndr.* 2011; 57(3): 190-6.
  28. Kling HM, Shipley TW, Patil SP, Kristoff J, Bryan M, Montelaro RC, et al. Relationship of *Pneumocystis jirovecii* humoral immunity to prevention of colonization and chronic obstructive pulmonary disease in a primate model of HIV infection. *Infect Immun.* 2010; 78(10): 4320-30.
  29. Wells J, Haidaris CG, Wright TW, Gigliotti F. Active immunization against *Pneumocystis carinii* with a recombinant P. carinii antigen. *Infect Immun.* 2006; 74(4): 2446-8.
  30. Stasaka S, Kobayashi S, Yagi K, Asami T, Namkoong H, Yamasawa W, et al. erum (1 → 3) β-d-glucan assay for discrimination between *Pneumocystis jirovecii* pneumonia and colonization. *J Infect Chemother.* 2014; 24: 1341-321.
  31. Huang SN, Fischer SH, O'Shaughnessy E, Gill VJ, Masur H, Kovacs JA. Development of a PCR assay for diagnosis of *Pneumocystis carinii* pneumonia based on amplification of the multicopy major surface glycoprotein gene family. *Diagn Microbiol Infect Dis.* 1999; 35(1): 27-32.
  32. Matsumura Y, Ito Y, Yamamoto M, Matsushima A, Nagao M, Takakura S, et al. *Pneumocystis* polymerase chain reaction and blood (1→3)-β-D-glucan assays to predict survival with suspected *Pneumocystis jirovecii* pneumonia. *J Infect Chemother.* 2014; 20(2): 109-14
  33. Fischl MA, Dickinson GM, La Vole L. Safety and efficacy of sulfamethoxazole and trimethoprim chemoprophylaxis for *Pneumocystis carinii* pneumonia in AIDS. *JAMA.* 1988; 259(8): 1185-9.
  34. Hardy WD, Feinberg J, Finkelstein DM, Power ME, He W, Kaczka C, et al. A controlled trial of trimethoprim-sulfamethoxazole or aerosolized pentamidine for secondary prophylaxis of *Pneumocystis carinii* pneumonia in patients with the acquired immunodeficiency syndrome: AIDS Clinical Trials Group protocol 021. *N Engl J Med.* 1992; 327(26): 1842-8.
  35. Huang L, Morris AM, Beard CB. *Pneumocystis carinii* hydroxyterate synthase mutations and treatment with sulfa or sulfone regimens: a proposal for standardized definitions for clinical evaluation. *J Eukaryot Microbiol.* 2001; 48: 180-181.
  36. Toma E, Fournier S, Dumont M, Bolduc P, Deschamps H. Clindamycin/primaquine versus trimethoprim-sulfamethoxazole as primary therapy for *Pneumocystis carinii* pneumonia in AIDS: a randomized, double-blind pilot trial. *Clin Infect Dis.* 1993; 17(2): 178-84.
  37. Leoung GS, Mills J, Hopewell PC, Hughes W, Wofsy C. Dapsone-trimethoprim for *Pneumocystis carinii* pneumonia in the acquired immunodeficiency syndrome. *Ann Intern Med.* 1986; 105(1): 45-8.
  38. Safrin S, Finkelstein DM, Feinberg J, Frame P, Simpson G, Wu A, et al. Comparison of three regimens for treatment of mild to moderate *Pneumocystis carinii* pneumonia in patients with AIDS: A double-blind, randomized trial of oral trimethoprim-sulfamethoxazole, dapsone-trimethoprim, and clindamycin-primaquine. *Ann Intern Med.* 1996; 124(9): 792-802.
  39. Dohn MN, Weinberg WG, Torres RA, Follansbee SE, Caldwell PT, Scott JD, et al. Oral atovaquone compared with intravenous pentamidine for *Pneumocystis carinii* pneumonia in patients with AIDS. *Ann Intern Med.* 1994; 121(3): 174-80.
  40. Rosenberg DM, McCarthy W, Slavinsky J, Chan CK, Montaner J, Braun J, et al. Atovaquone suspension for treatment of *Pneumocystis carinii* pneumonia in HIV-infected patients. *AIDS.* 2001; 15(2): 211-4.
  41. Sattler FR, Frame P, Davis R, Nichols L, Shelton B, Akil B, et al. Trimetrexate with leucovorin versus trimethoprim-sulfamethoxazole for moderate to severe episodes of *Pneumocystis carinii* pneumonia in patients with AIDS: a prospective, controlled multicenter investigation of the AIDS Clinical Trials Group Protocol 029/031. *J Infect Dis.* 1994; 170(1): 165-72.
  42. Procop GW, Haddad S, Quinn J, Wilson ML, Henshaw NG, Reller LB, et al. Detection of *Pneumocystis jirovecii* in respiratory specimens by four staining methods. *J Clin Microbiol* 2004; 42: 3333-5.
  43. Kaplan JE, Benson C, Holmes KK, Brooks JT, Pau A, Masur H. Guidelines for prevention and treatment of opportunistic infections in HIV-infected adults and adolescents. *MMWR Recomm Rep.* 2009; 58(4): 1-207.
  44. Klein NC, Duncanson FP, Lenox TH, Forszpaniak C, Sherer CB, Quentzel H, et al. Trimethoprim-sulfamethoxazole versus pentamidine for *Pneumocystis carinii* pneumonia in AIDS patients: results of a large prospective randomized treatment trial. *AIDS.* 1992; 6(3):301-5.
  45. Conte JE, Hollander H, Golden JA. Inhaled or Reduced-Dose Intravenous Pentamidine for *Pneumocystis carinii* Pneumonia A Pilot Study. *Ann Intern Med.* 1987; 107(4): 495-8.
  46. Golden J, Hollander H, Chernoff D, Feigal D, Conte J. Prevention of *Pneumocystis carinii* pneumonia by inhaled pentamidine. *Lancet.* 198; 333(8639): 654-7.
  47. Sattler FR, Cowan R, Nielsen DM, Ruskin J. Trimethoprim-Sulfamethoxazole Compared with

- Pentamidine for Treatment of *Pneumocystis carinii* Pneumonia in the Acquired Immunodeficiency Syndrome A Prospective, Noncrossover Study. *Ann Intern Med.* 1988; 109(4): 280-7.
48. Smith RM, Iwamoto GK, Richerson HB, Flaherty JP. Trimethoprim-sulfamethoxazole desensitization in the acquired immunodeficiency syndrome. *Ann Intern Med.* 1987; 106(2): 335.
49. Kazanjian P, Armstrong W, Hossler PA, Burman W, Richardson J, Lee C-H, et al. *Pneumocystis carinii* mutations are associated with duration of sulfa or sulfone prophylaxis exposure in AIDS patients. *J Infect Dis.* 2000; 182(2): 551-7.
50. Kazanjian P, Locke AB, Hossler PA, Lane BR, Bartlett MS, Smith JW, et al. *Pneumocystis carinii* mutations associated with sulfa and sulfone prophylaxis failures in AIDS patients. *AIDS.* 1998; 12(8): 873-8.
51. Ma L, Borio L, Masur H, Kovacs JA. *Pneumocystis carinii* dihydropteroate synthase but not dihydrofolatereductase gene mutations correlate with prior trimethoprim-sulfamethoxazole or dapsone use. *J Infect Dis.* 1999; 180(6): 1969-78.
52. Helweg-Larsen J, Benfield TL, Eugen-Olsen J, Lundgren JD, Lundgren B. Effects of mutations in *Pneumocystis carinii* dihydropteroate synthase gene on outcome of AIDS-associated *P carinii* pneumonia. *Lancet.* 1999; 354(9187): 1347-51.
53. Cushion MT, Linke MJ, Ashbaugh A, Sesterhenn T, Collins MS, Lynch K, et al. Echinocandin treatment of pneumocystis pneumonia in rodent models depletes cysts leaving trophic burdens that cannot transmit the infection. *PLoS One.* 2010; 5(1): e8524.
54. Deresinski SC, Stevens DA. Caspofungin. *Clin Infect Dis.* 2003; 36(11): 1445-57.